

Karyotypic features including organizations of the 5S, 45S rDNA loci and telomeres of *Scadoxus multiflorus* (Amaryllidaceae)

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Academic editor: L. Peruzzi | Received 20 July 2016 | Accepted 7 October 2016 | Published 22 November 2016

<http://zoobank.org/573FB0FF-22E8-412E-A853-D46BD88B3F97>

Citation: Monkheang P, Chaveerach A, Sudmoon R, Tanee T (2016) Karyotypic features including organizations of the 5S, 45S rDNA loci and telomeres of *Scadoxus multiflorus* (Amaryllidaceae). Comparative Cytogenetics 10(4): 637–646. doi: 10.3897/CompCytogen.v10i4.9958

Abstract

Scadoxus multiflorus Martyn, 1795 is an ornamental plant with brilliantly colored flowers. Even though its chromosomes are rather large, there is no karyotype description reported so far. Therefore, conventional and molecular cytogenetic studies including fluorescence *in situ* hybridization (FISH) with 45S and 5S rDNA, and human telomere sequence (TTAGGG)_n probes (Arabidopsis-type telomere probes yielded negative results) were carried out. The chromosome number is as reported previously, 2n = 18. The nine chromosome pairs include two large submetacentric, five large acrocentric, one medium acrocentric, two small metacentric and eight small submetacentric chromosomes. Hybridization sites of the 45S rDNA signals were on the short arm ends of chromosomes #1, #3 and #8, while 5S rDNA signals appeared on the long arm of chromosome 3, in one homologue as a double signal. The telomere signals were restricted to all chromosome ends. Three chromosome pairs could be newly identified, chromosome pair 3 by 5S rDNA and chromosomes #1, #3 and #8 by 45S rDNA loci. In addition to new information about rDNA locations we show that the ends of *S. multiflorus* chromosomes harbor human instead of Arabidopsis-type telomere sequences. Overall, the *S. multiflorus* karyotype presents chromosomal heteromorphy concerning size, shape and 45S and 5S rDNA positioning. As *Scadoxus* Rafinesque, 1838 and related species are poorly studied on chromosomal level the here presented data is important for better understanding of evolution in Amaryllidaceae.

Keywords

Scadoxus multiflorus, aceto-orcein staining, fluorescence *in situ* hybridization (FISH), rDNA, telomere

Introduction

Scadoxus multiflorus Martyn, 1795 (also known as *Haemanthus multiflorus* Martyn, 1795 and *H. kalbreyeri* Baker, 1878) is a species belonging to the family Amaryllidaceae (Chase et al. 2009), which can naturally only be found in Southern and tropical Africa (Patwary and Zaman 1980). Formerly, the genus *Scadoxus* Rafinesque, 1838 was *Haemanthus* Linnaeus, 1753 and was included in the family Liliaceae (Ahirwar and Verma 2014); however, it is not included in the current circumscription of this family (Peruzzi 2016), and was indeed separated out and relocated by Angiosperm Phylogeny Group (APG IV) classification system.

Scadoxus multiflorus is economically important as it is popular as cultivated ornamental plant. Nonetheless, this species was not studied yet for its karyotype in details. There are only two previous reports based on conventional chromosome staining of *S. multiflorus*. Both studies reported $2n = 18$ chromosomes, however, Ahirwar and Verma (2014) found six submetacentric and 12 acrocentric chromosomes while Patwary and Zaman (1980) reported one metacentric, nine submetacentric, six acrocentric and two telocentric chromosomes. Besides these contradictory data, molecular cytogenetic approaches like fluorescence *in situ* hybridization (FISH) have not been applied in this species yet. Such methods being available since the late 1980s (Li et al. 2016, Taguchi et al. 2016) enable detection, characterization and localization of rDNA regions (Chirino et al. 2015) and/or telomeres. The latter are known to be important to protect chromosomal ends of all eukaryotes against nucleolytic degradation, non-homologous end-joining and replication-mediated shortening. They usually consist of short tandem repeats, such as $(TTTAGGG)_n$ in *Arabidopsis* Heynhold, 1842 (Richards and Ausubel 1988) or $(TTAGGG)_n$ in humans (Moyzis et al. 1988). The *Arabidopsis* G-rich telomere repeat is rather conserved and has been detected at the ends of most chromosomes of higher plants examined so far. Nevertheless, in some plants the TTTAGGG-type telomere repeat is lacking and substituted by other repeat sequences (Fajkus et al. 2016).

As for *S. multiflorus* reported karyotypic details are contradictory and no FISH studies have been performed so far the present study aimed to close this gap of knowledge using conventional staining and FISH.

Material and methods**Plant material and chromosome preparation**

Scadoxus multiflorus plant was collected in Khon Kaen Province, Northeastern, Thailand (A. Chaveerach et al. 903, Department of Biology, Faculty of Science, Khon Kaen Univer-

sity, Khon Kaen, Thailand). The roots were collected from bulbs placed in distilled water at room temperature. The root tips were excised and kept in cold water for 1 h at 4°C, after that transferred to 0.05% colchicine solution for 4 h at room temperature to accumulate metaphase chromosomes before fixation in ethanol:acetic acid (3:1, v/v) for at least 24 h at 4°C. The protocol for the SteamDrop method (Kirov et al. 2014) was adopted with a few modifications. Briefly, fixed root tips were washed twice with enzyme buffer (0.01 M citric acid, 0.01 M sodium citrate, pH 4.7) to remove the fixative and digested at 37°C for 4 h in enzyme solution consisting of 0.7% cellulase R10 (Duchefa C8001), 0.7% cellulase (CalBioChem 319466), 1% pectolyase (Sigma P3026) and 1% cytohellicase (Sigma C8274) in enzyme buffer. Then, the soft meristematic tissues were washed twice with distilled water and 96% ethanol to remove supernatant with centrifuge before broken with a dissecting needle in a tube in fixative. The suspension was dropped on a glass slide and air dried. Preparations with well spread metaphases were selected for further analyses.

Orcein staining and idiogram generation

Conventional staining was carried out on slides using 2% aceto-orcein for 5 min at room temperature and then covered with a coverslip. Ten well-spread metaphases were selected for photomicrography with a digital camera under oil immersion by light microscope. The length of short and long chromosome arms (p and q) were measured separately and added to calculate the total length (LT). The relative length of chromosome (RL), the centromeric index (CI) and standard deviation (SD) of RL and CI were calculated according to Chaivasut (1989). The CI ($q/p+q$) between 0.500–0.599, 0.600–0.699, 0.700–0.899 and 0.900–0.999 were considered as metacentric (m), submetacentric (sm), acrocentric (a) and telocentric (t), respectively, following Turpin and Lejeune (1965) to classify the types of chromosome. These parameters were used for idiogramming by computer. Comparison of different estimators of intrachromosomal asymmetry was performed, including mean centromeric asymmetry (M_{CA}) and coefficient of variation of chromosome length (CV_{CL}) based on the equations provided by Peruzzi and Eroğlu (2013). Metaphase chromosomes from overall 10 cells were included.

DNA probe generation and labeling

45S and 5S rDNAs which were isolated from *Arabidopsis thaliana* Schur, 1866 and telomere repeat sequences from *Arabidopsis* (TTTAGGG)_n and human (TTAGGG)_n were applied in this study. The plasmid of 45S rDNA cloned in the vector T₁₅P₁₀IV₁ was labelled with Alexa 488-dUTP, while the 5S rDNA probe was labelled with Cy3-dUTP by Nick translation (Roche Cat No 11745808910). The telomeric probes were generated by polymerase chain reaction (PCR) in absence of a DNA template using primers (TTTAGGG)₅ and (CCCTAAA)₅, and (TTAGGG)₅ and (CCCTAA)₅ according to Ijdo et al. (1991) and labelled with Cy3-dUTP by nick translation.

Fluorescence *in situ* hybridization

A FISH protocol according to Lysak et al. (2006) was applied with minor modifications. The slide with fixed metaphase cells was washed in $2 \times$ SSC (300 mM Na-citrate, 30 mM NaCl, pH 7.0) for 5 min at room temperature and treated with 45% acetic acid for 3 min. Then the slides were washed twice in $2 \times$ SSC for 5 min each at room temperature before digestion in pepsin solution (10 mg/ml) in 0.01 M HCl for 1 min at 37°C, rinsing twice in $2 \times$ SSC for 5 min, post-fixation in 4% formaldehyde in $2 \times$ SSC for 5 min at room temperature, and two washes in $2 \times$ SSC, with final dehydration in an ethanol series (70%, 90%, 100%) for 2 min, each, at room temperature and air drying.

A 3 μ l (60 ng) of labelled probe was dissolved in 17 μ l of hybridization mixture (20% dextran sulfate, 50% formamide in $2 \times$ SSC, pH 7.0), and pre-denatured at 95°C for 5 min. Then the solution was added to the slide, covered with a coverslip and sealed by rubber gum. Now, the slide was placed on a heating plate at 80°C for 2 min for co-denaturation of probe and target DNA and incubated in a moist chamber at 37°C for 18 h for hybridization.

After hybridization, slides were washed in $2 \times$ SSC for 5 min at 42°C and three times with 50% formamide in $2 \times$ SSC for 5 min at 42°C. After that, slides were washed three times with $2 \times$ SSC for 5 min at 42°C for 5 min each. Finally, the slides were dehydrated in an ethanol series, air dried, counterstained with 4, 6-diamino-2-phenylindole (DAPI) plus Vectashield antifade mounting medium (Vector Laboratories, USA) and covered with a coverslip. Signals were detected using an epifluorescence microscope with Triple filter (UV, Texas Red and FITC) and photographed (microscope: Axioplan2, Zeiss; Camera: Hammamatsu-ORCA-ER C4742-80, Japan; Lamp: Flouarc, Zeiss).

Results

The idiogram and karyotype analyses established from the metaphases confirmed the diploid chromosome number of *S. multiflorus* to be $2n = 18$. The karyotype analysis of this species is summarized in Table 1. The range of total arm length, centromeric index and relative chromosome length are 3.78–16.01 μ m, 0.57–0.87 μ m and 0.04–0.15 μ m respectively. The karyotype comprises two large submetacentric, five large acrocentric, one medium acrocentric, two small metacentric and eight small submetacentric chromosomes (Fig. 1). The plant shows a clear tendency to have karyotypes distinct on asymmetry grounds with relatively low M_{CA} and CV_{CL} as shown in Figure 2 and Table 2.

The metaphase spreads were hybridized with 45S (Alexa, green) and 5S (Cy3, red) rDNA probes as shown in Fig. 3. The hybridization signals for the 45S rDNA probe are in terminal positions of the short arms of chromosomes #1, #3 and #8 (Fig. 3A). The 5S rDNA signals were detected on the long arms of chromosome #3 with one homologue showing two adjacent signals (Figs. 3B–C).

Table 1. Mean length of the short arm chromosome (Ls), long arm chromosome (Ll), total arm chromosome (LT), centromeric index (CI), relative length (RL) and standard deviation (SD) of CI, RL from metaphase chromosomes in 10 cells of the blood lily (*Scadoxus multiflorus*), 2n = 18.

Chr.	Ls	Ll	LT	CI \pm SD	RL \pm SD	Type	Size
1a*	4.86	11.14	16.007	0.696 \pm 0.068	0.148 \pm 0.010	sm	L
1b*	4.66	9.81	14.465	0.678 \pm 0.100	0.134 \pm 0.025	sm	L
2a*	2.85	9.89	12.741	0.776 \pm 0.085	0.118 \pm 0.008	a	L
2b*	2.86	9.00	11.865	0.759 \pm 0.073	0.110 \pm 0.015	a	L
3	1.49	9.52	11.008	0.865 \pm 0.131	0.102 \pm 0.015	a	L
4a*	2.29	8.58	10.870	0.790 \pm 0.087	0.101 \pm 0.013	a	L
4b*	2.15	7.33	9.480	0.773 \pm 0.074	0.088 \pm 0.016	a	M
5	1.62	3.17	4.784	0.662 \pm 0.066	0.044 \pm 0.006	sm	S
6	1.95	2.53	4.485	0.565 \pm 0.034	0.042 \pm 0.004	m	S
7	1.50	2.90	4.402	0.659 \pm 0.063	0.041 \pm 0.004	sm	S
8	1.50	2.65	4.153	0.639 \pm 0.065	0.038 \pm 0.004	sm	S
9	1.19	2.59	3.779	0.685 \pm 0.069	0.035 \pm 0.005	sm	S

Remarks: Chr. = chromosome pair, a = acrocentric, m = metacentric, sm = submetacentric, L = large, M = medium, S = small, a* and b* = heteromorphic pairs 1, 2, and 4.

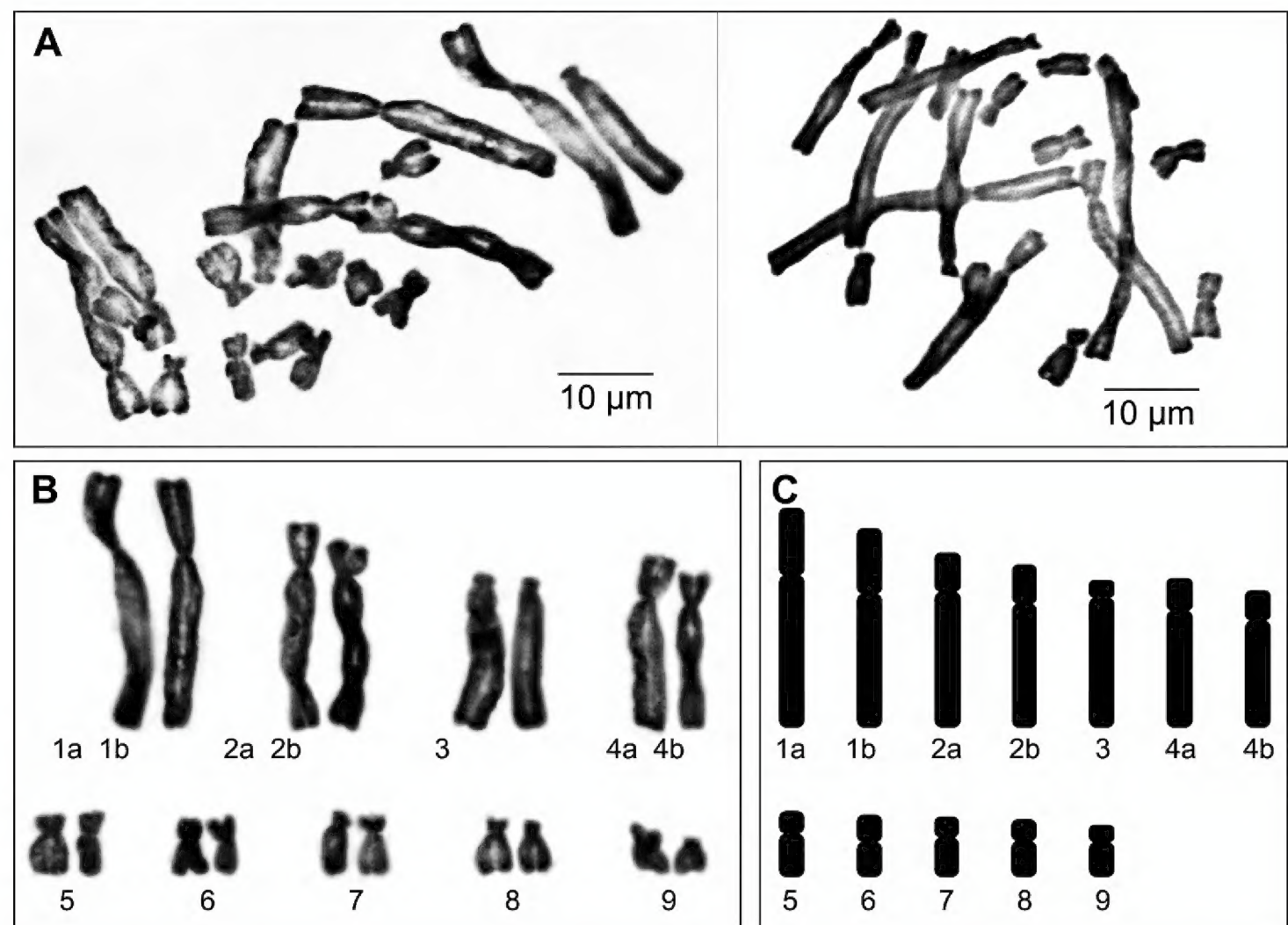


Figure 1. Mitotic metaphase chromosomes of *Scadoxus multiflorus* (2n = 18). **A** aceto-orcein staining of two cells **B** karyogram showing four large and five small pairs of chromosomes **C** idiogram.

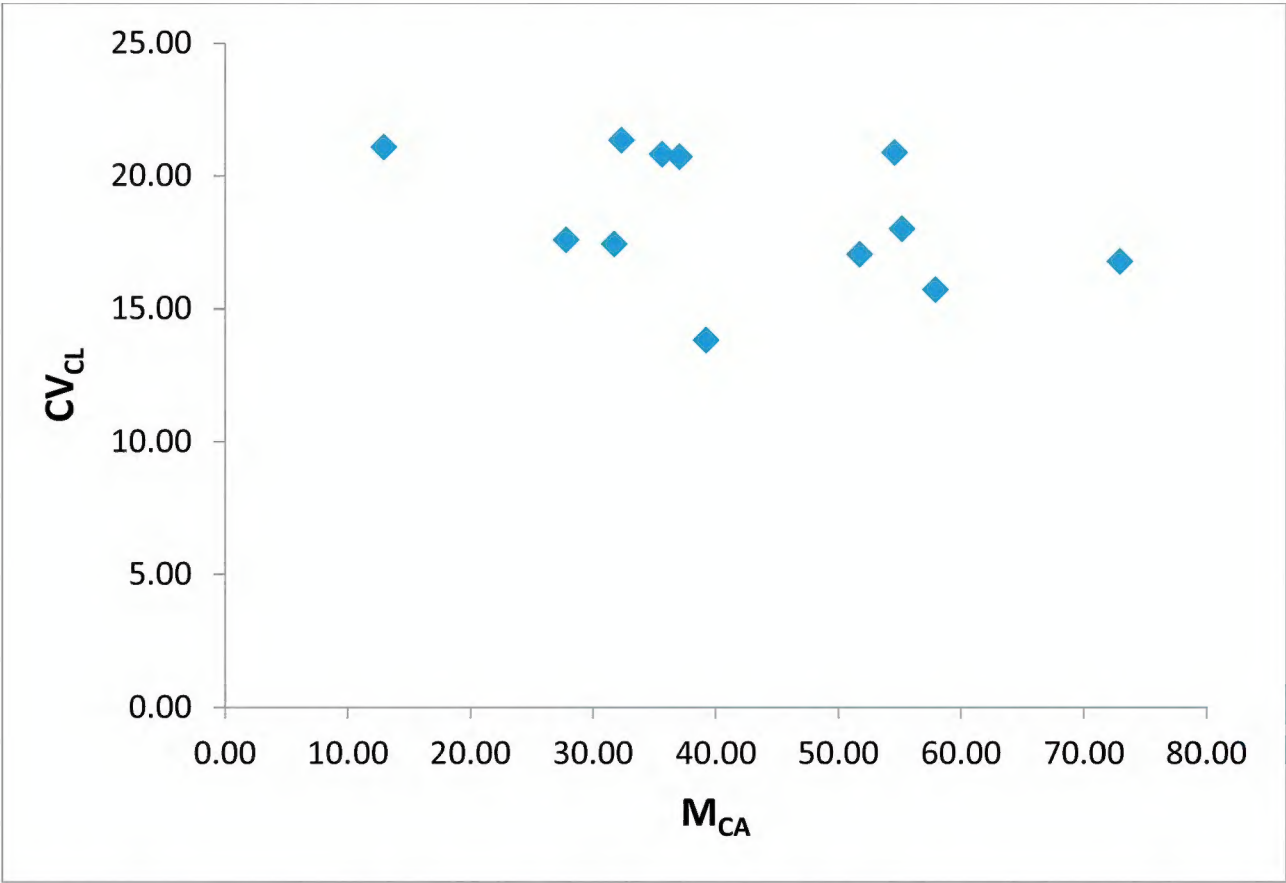


Figure 2. Scatter plot of M_{CA} against CV_{CL} of *Scadoxus multiflorus* chromosomes.

Table 2. Comparison of different estimators of intrachromosomal asymmetry including mean centromeric asymmetry (M_{CA}) and coefficient of variation of chromosome length (CV_{CL}) from metaphase chromosomes in 10 cells of the blood lily (*Scadoxus multiflorus*), $2n = 18$.

Chr.	Mean from 10 metaphases						SD of chr. length	M_{CA}	CV_{CL}
	Ll-Ls	Ls Ll	Ls (Ll+Ls)	Ll (Ll+Ls)	(Ll-Ls) Ll	(Ll-Ls) (Ll+Ls)			
1a*	6.28	0.44	0.30	0.70	0.56	0.39	2.21	39.23	13.82
1b*	5.15	0.47	0.32	0.68	0.53	0.36	3.01	35.61	20.82
2a*	7.03	0.29	0.22	0.78	0.71	0.55	2.30	55.20	18.02
2b*	6.14	0.32	0.24	0.76	0.68	0.52	2.02	51.72	17.05
3	8.03	0.16	0.14	0.86	0.84	0.73	1.85	72.94	16.78
4a*	6.29	0.27	0.21	0.79	0.73	0.58	1.71	57.90	15.71
4b*	5.18	0.29	0.23	0.77	0.71	0.55	1.98	54.60	20.89
5	1.55	0.51	0.34	0.66	0.49	0.32	1.02	32.34	21.35
6	1.40	0.52	0.34	0.66	0.48	0.32	0.77	31.74	17.44
7	0.58	0.77	0.44	0.56	0.23	0.13	0.95	12.93	21.09
8	1.15	0.57	0.36	0.64	0.43	0.28	0.73	27.79	17.60
9	1.40	0.46	0.31	0.69	0.54	0.37	0.78	37.04	20.73

Remarks: Chr. = chromosome pair, a = acrocentric, m = metacentric, sm = submetacentric, L = large, M = medium, S = small, a* and b* = heteromorphic pairs 1, 2, and 4.

The hybridization of the Arabidopsis-type telomeric probe yielded no obvious FISH signal, while the human-type (Cy3, red) revealed small signals exclusively at the end of all *S. multiflorus* chromosomes (Fig. 4).

Overall, the results show chromosomal heteromorphy in sizes and shapes.

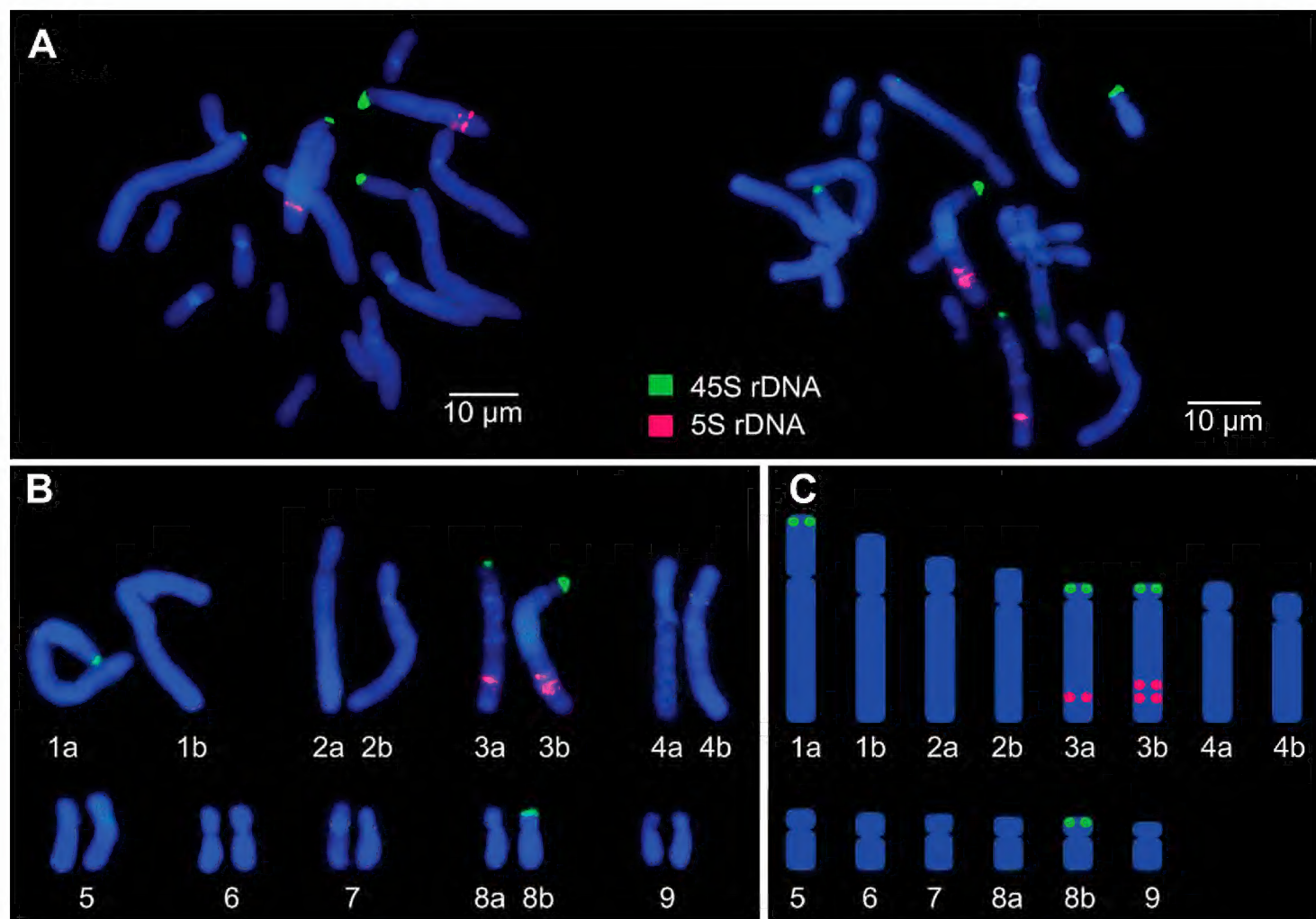


Figure 3. Organizations of 45S rDNA and 5S rDNA loci on metaphase chromosomes of *Scadoxus multiflorus* ($2n = 18$). **A** FISH signals of 45S (Alexa, green) and 5S (Cy3, red) rDNA probes on two cells **B** karyogram **C** idiogram.

Discussion

Here we provide the first study of *S. multiflorus* chromosomes by means of molecular cytogenetics. Furthermore, the previously reported chromosome number could be confirmed to be $2n = 18$ (Patwary and Zaman 1980, Ahirwar and Verma 2014). However, here we report a karyotype variant in *S. multiflorus* as carrying two metacentric, ten submetacentric and six acrocentric chromosomes, which is in contradiction to what was previously reported by Patwary and Zaman (1980) or Ahirwar and Verma (2014). This result indicates that further studies are necessary to clarify if there are either (i) cryptic (sub-)species in *S. multiflorus*, (ii) assessment problems in any of the previous studies, or (iii) any kind of chromosomal heteromorphisms leading to the observed different chromosomal shapes.

Although the regions of 5S and 45S were not observed with conventional staining, due to several limitations, such as oil-immersion light microscopic methods, genetic processing and analysis standards, the signal intensities of rDNA probes in FISH showed clear variation in copy numbers. The copy number and distributed position on chromosome are very important as species markers (Boocock et al. 2015, Li et al 2016).

In a group of families of the monocot order Asparagales, the telomeric sequence (TTAGGG)_n of the human-type was found to be maintained (Sykorova et al. 2003).

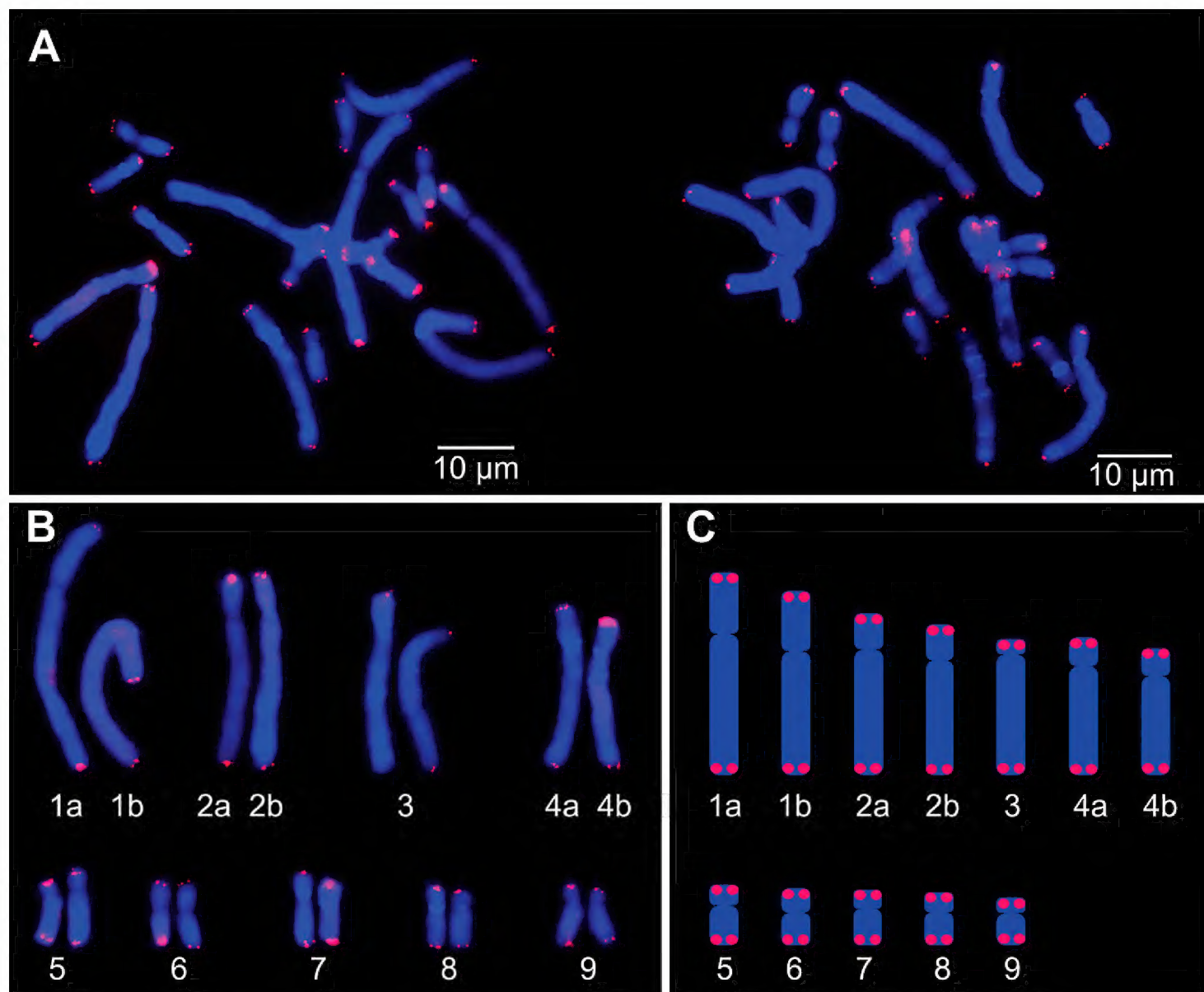


Figure 4. Localization of the human telomere repeat sequence (TTAGGG)_n on metaphase chromosomes of *Scadoxus multiflorus* (2n = 18). **A** FISH signals of the (TTAGGG)_n probe on two cells **B** karyogram **C** idiogram.

Thus it is in accordance with the literature that *S. multiflorus* also has this type of telomeric repeat, as the family Amaryllidaceae belongs to this branch of Asparagales which switched from *Arabidopsis*-type to human type telomere sequence. The knowledge is the one factor supports the classification theory of a common ancestor for a plant group.

Overall, our results allow now distinguishing five of the nine *S. multiflorus* chromosome pairs individually. Development of suitable genomic single-copy FISH probes might allow discrimination of all chromosome pairs and to use them for identification of homologous chromosomes in other species of genus, or even of related genera. As *Scadoxus* and related species are poorly studied on chromosomal level the here presented data is important for better understanding of evolution in Amaryllidaceae.

Acknowledgements

The authors thanks to Ingo Schubert, for providing plant materials and labelled probes, and all members of the Quantitative Genetics and the Chromosome Structure and

Function Research Group of the Leibniz Institute of Plant Genetics and Crop Plant Research (IPK), Germany for offering laboratory facilities and assistance. Thanks to the Research and Study in Abroad Fund for Graduated Students from Faculty of Science, Khon Kaen University for providing financial support to research in Germany. We are thankful for critical comments and helpful suggestions by Thomas Liehr, Institute of Human Genetics, Universitätsklinikum Jena, Germany.

References

- Ahirwar R, Verma RC (2014) Karyotypic studies in some members of Liliaceae. *The Journal of Cytology and Genetics* 15(NS): 61–74. <http://socg.in/jrnls/2014/part2.pdf>
- Boocock J, Chagné D, Merriman TR, Black MA (2015) The distribution and impact of common copy-number variation in the genome of the domesticated apple, *Malus x domestica* Borkh. *BMC Genomics* 16: 848. doi: 10.1186/s12864-015-2096-x
- Chaiyasut K (1989) Cytogenetic and Cytotaxonomy of the Family *Zephyranthes*. Department of Botany, Faculty of Science, Chulalongkorn University, Bangkok.
- Chase MW, Reveal JL, Fay MF (2009) A subfamilial classification for the expanded asparagalean families Amaryllidaceae, Asparagaceae and Xanthorrhoeaceae. *Botanical Journal of the Linnean Society* 161(2): 132–136. doi: 10.1111/j.1095-8339.2009.00999.x
- Chirino MG, Rossi LF, Bressa MJ, Luaces JP, Merani MS (2015) Comparative study of mitotic chromosomes in two blowflies, *Lucilia sericata* and *L. cluvia* (Diptera, Calliphoridae), by C- and G-like banding patterns and rRNA loci, and implications for karyotype evolution. *Comparative Cytogenetics* 9(1): 103–118. doi: 10.3897/CompCytogen.v9i1.8671
- Fajkus P, Peška V, Sitová Z, Fulnečková J, Dvořáčková M, Gogela R, Sýkorová E, Hapala J, Fajkus J (2016) *Allium* telomeres unmasked: the unusual telomeric sequence (CTCGGT-TATGGG)_n is synthesized by telomerase. *Plant Journal* 85(3): 337–347. doi: 10.1111/tpj.13115
- Ijdo JW, Wells RA, Baldini A, Reeders ST (1991) Improved telomere detection using a telomere repeat probe (TTAGGG)_n generated by PCR. *Nucleic Acids Research* 19(17): 4780. doi: 10.1093/nar/19.17.4780
- Kirov I, Divashuk M, Laere KV, Soloviev A, Khrustaleva L (2014) An easy “SteamDrop” method for high quality plant chromosome preparation. *Molecular Cytogenetics* 7(21): 1–10. doi: 10.1186/1755-8166-7-21
- Li X, Yang Z, Liao H, Zhang Z, Huang X, Bao Z (2016) Chromosomal mapping of tandem repeats in the Yesso Scallop, *Patinopecten yessoensis* (Jay, 1857), utilizing fluorescence *in situ* hybridization. *Comparative Cytogenetics* 10(1): 157–169. doi: 10.3897/CompCytogen.v10i1.7391
- Lysak MA, Fransz P, Schubert I (2006) Cytogenetic analyses of *Arabidopsis*. *Methods in Molecular Biology* 323: 173–186. doi: 10.1385/1-59745-003-0:173
- Moyzis RK, Buckingham JM, Crams LS, Dani M, Deaven LL, Jones MD, Meyne J, Ratliff RL, Wu JR (1988) A highly conserved repetitive DNA sequence, (TTAGGG)_n, present at the telomeres of human chromosomes. *Proceedings of the National Academy of Sciences of*

- the United States of America 85: 6622–6626. <http://www.pnas.org/content/85/18/6622.long>
- Patwary MU, Zaman MA (1980) Cytogenetic of Amaryllidaceae II. Heterozygosity in *Haemanthus kalbreyeri* Baker. *Caryologia* 33: 461–468. doi: 10.1080/00087114.1980.10796860
- Peruzzi L, Eroğlu HE (2013) Karyotype asymmetry: again, how to measure and what to measure. *Comparative Cytogenetics* 7: 1–9. doi: 10.3897/CompCytogen.v7i1.4431
- Peruzzi L (2016) A new infrafamilial taxonomic setting for Liliaceae, with a key to genera and tribes. *Plant Biosyst* 150: 1341–1347. doi: 10.1080/11263504.2015.1115435
- Richards E, Ausubel F (1988) Isolation of a higher eukaryotic telomere from *Arabidopsis thaliana*. *Cell* 53: 127–136. doi: 10.1016/0092-8674(88)90494-1
- Sykorova E, Lim KY, Kunicka Z, Chase MW, Bennett MD, Fajkus J, Leitch AR (2003) Telomere variability in the monocotyledonous plant order Asparagales. *Proceedings of the Royal Society B* 270: 1893–1904. doi: 10.1098/rspb.2003.2446
- Taguchi T, Kubota S, Mezaki T, Tagami E, Sekida S, Nakachi S, Okuda K, Tominaga A (2016) Identification of homogeneously staining regions by G-banding and chromosome microdissection, and FISH marker selection using human Alu sequence primers in a scleractinian coral *Coelastrea aspera* Verrill, 1866 (Cnidaria). *Comparative Cytogenetics* 10(1): 61–75. doi: 10.3897/CompCytogen.v10i1.5699
- Turpin R, Lejeune J (1965) Les chromosome humains (Caryotype Nomal et Variations Pathologiques). Gauthier-Villars, Paris.